

Drought Induced Biochemical Changes in *Commiphora wightii*

Vineet Soni^{1*}, P. L. Swarnkar²

¹Plant Bioenergetics and Biotechnology Laboratory, Department of Botany, Mohanlal Sukhadia University, Udaipur, India,

²Department of Botany, University of Rajasthan, Jaipur, India

*Address for Correspondence: Dr. Vineet Soni, Assistant Professor, Department of Botany, Mohanlal Sukhadia University, Udaipur, India

Received: 18 March 2017/Revised: 28 May 2017/Accepted: 23 June 2017

ABSTRACT- The effects of water deficit induced by withholding water in soil pots were examined on the activities of various key enzymes i.e. nitrate reductase, peroxidase, acid phosphatase, α -amylase and invertase in *Commiphora wightii*. Drought stress induced decrease in the activities of nitrate reductase, peroxidase, α -amylase and invertase was observed in leaves of *C. wightii*. The decreased activity of peroxidase enzyme in *C. wightii* plants under water stress condition indicates that the plant is capable of maintaining growth vigour despite adverse conditions. On the other hand, acid phosphatase activity increased continuously in the leaves of *C. wightii* plants subjected to water stress. The results clearly indicate that regulation of enzymatic activity under drought is an essential biochemical process, which prevents the plants from drought-induced damage.

Key-words- Drought, *Commiphora wightii*, Nitrate reductase, Peroxidase, Acid phosphatase, α -amylase, Invertase

INTRODUCTION

Drought is one of the important biomass-limiting stress factors which affect practically every aspect of plant growth and metabolism [1]. Plant's responses to water deficit condition depend upon various factors such as duration and degree of stress, growth stage and time of stress exposure. During drought the water potential (ψ), relative water content (RWC) and net photosynthetic CO₂ fixation (A) significantly decrease [2]. The stomatal closure is among the first responses to the water stress, and is assumed to be the main cause of impaired photosynthesis, since the stomatal closure limits CO₂ availability to the mesophyll. On the other hand, the limitation of CO₂ fixation during drought is also influenced by the diffusion of CO₂ from the intercellular spaces to chloroplasts [3], and by other metabolic factors such as changes in the activity of ribulose-1,5-bisphosphate-carboxylase and perturbed regeneration of ribulose-1,5-bisphosphate etc [3].

C. wightii (Arnott) Bhandari (Burseraceae) is a slow growing, highly branched, and critically endangered plant that grows wild in the arid and semi-arid regions of India, Pakistan and tropical regions of Africa.

In India, the plant is widespread in the dry and arid region of Rajasthan and Gujarat states. Plants grown in such arid and semi-arid regions often encounter various stress conditions such as high temperature, high wind regime, high light intensity, drought etc. Against drought, plants adapt themselves by different mechanisms, including change in morphological and developmental pattern as well as physiological and biochemical responses [4]. Adaptation to drought is associated with metabolic adjustments that lead to the modulation of different enzymes. It has been hypothesized that these particular changes induced under water deficit conditions enable the plant to endure drought stress. Therefore, in the present investigations, efforts were carried out to understand the biochemical basis of drought tolerance in *C. wightii* plants.

MATERIALS AND METHODS

Plant materials and Drought treatment- Two year old vegetatively propagated plants of *C. wightii* (Fig. 1 A) growing in pots at research nursery, Department of Botany, University of Rajasthan, Jaipur, India during March 2005 was used for the biochemical studies. The plants were divided into two sets (each of four plants), out of which one set was subjected to water stress by withholding of water supply till wilting symptoms appeared, while the second set was watered regularly and served as a control.

Sample preparation- Biochemical analyses of enzymes were carried out in the leaves as the leaves were devoid of resinous compounds, which interfere with biochemical

Access this article online

Quick Response Code



Website:

www.ijlssr.com

Crossref

DOI: 10.21276/ijlssr.2017.3.4.24

estimations. The samples were collected regularly at an interval of three days till day 15. The leaves were homogenized in pre-chilled mortar and pestle using appropriate buffers. The homogenate was centrifuged in high-speed centrifuge (KR 20000 T, KUBUTA, Japan) at 10,000 rpm for 20 minutes. The supernatant thus collected was used for enzyme assay and protein estimation. The results were averages of two replicates.

Enzyme assay- A method given by Jaworski [5] was used for assaying nitrate reductase activity. Guaiacol H_2O_2 method was used for assaying the activity of peroxidase [6]. The activity of acid phosphatase was assayed by using p-nitrophenyl phosphate as a substrate [7] and Bernfeld's method [8] was used for assaying the activity of α -amylase. A modified method of Harris and Jeffcoat [9] was used to determine the activity of acid invertase.

RESULTS AND DISCUSSION

Metabolic changes in response to water stress also include reduction in photosynthetic activity and accumulation of organic acids, such as malate, citrate and lactate accompanied by accumulation of proline, sugars and betaine [10,11]. Exposure of plants to low water potential often leads to loss of cell turgor and then plants undergo osmotic adjustments by the rapid accumulation of abscisic acid (ABA) and osmoprotectants [12]. In the present investigation, the activities of enzymes (nitrate reductase, peroxidase, acid phosphatase, α -amylase and invertase) and level of cellular metabolites (proteins and proline) were analyzed in *C. wightii* plants subjected to water deficit condition.

Nitrate reductase, which is involved in the reduction of NO_3^- to NO_2^- , is known to be highly sensitive to all types of stresses, particularly salt and drought stresses. In the

present study, a remarkable decline in the nitrate reductase

activity was observed after 3rd day of water deprivation (Fig. 1 B). On day 15 of water deprivation, the activity of nitrate reductase enzyme decreased to 15.6 % (± 2.02) of initial level. Similarly Inhibition of nitrate reductase under moisture stress has been reported by several workers in clusterbean [13], rice [14], corn [15], wheat [16], and Sorghum [17].

Peroxidase activity decreased continuously in the leaves of *C. wightii* plants subjected to water deficit conditions (Fig. 1 C). On day 15 the activity of this enzyme decreased to 30.7% (± 1.421) of initial level. The variation in peroxidase activity in the plants growing under normal irrigated condition was insignificant. The decrease in activity is ascribed to as a change in the structural proteins [18]. In general, the peroxidase activity is inversely correlated with the plant growth rates. The decreased activity of peroxidase enzyme in plants undergoes water stress condition indicates that the plant is capable of maintaining growth vigour despite adverse conditions.

Acid phosphatase is known to maintain a certain level of inorganic phosphate in plant cells under stress [19]. In the present study, acid phosphatase activity increased continuously in the leaves of *C. wightii* plants subjected to water stress (Fig. 1 D). Induction of acid phosphatase activity may be due to fact that under conditions of water stress, growth is restricted and delivery of phosphate is impaired, resulting in the activation of the cellular phosphatases that release soluble phosphate from its insoluble compounds inside or outside the cells [20]. Olmos and Hellin [21] observed that acid phosphatase under salt and water stress maintained a certain level of inorganic phosphate, which could be co-transported with H^+ along a gradient of proton motive force.

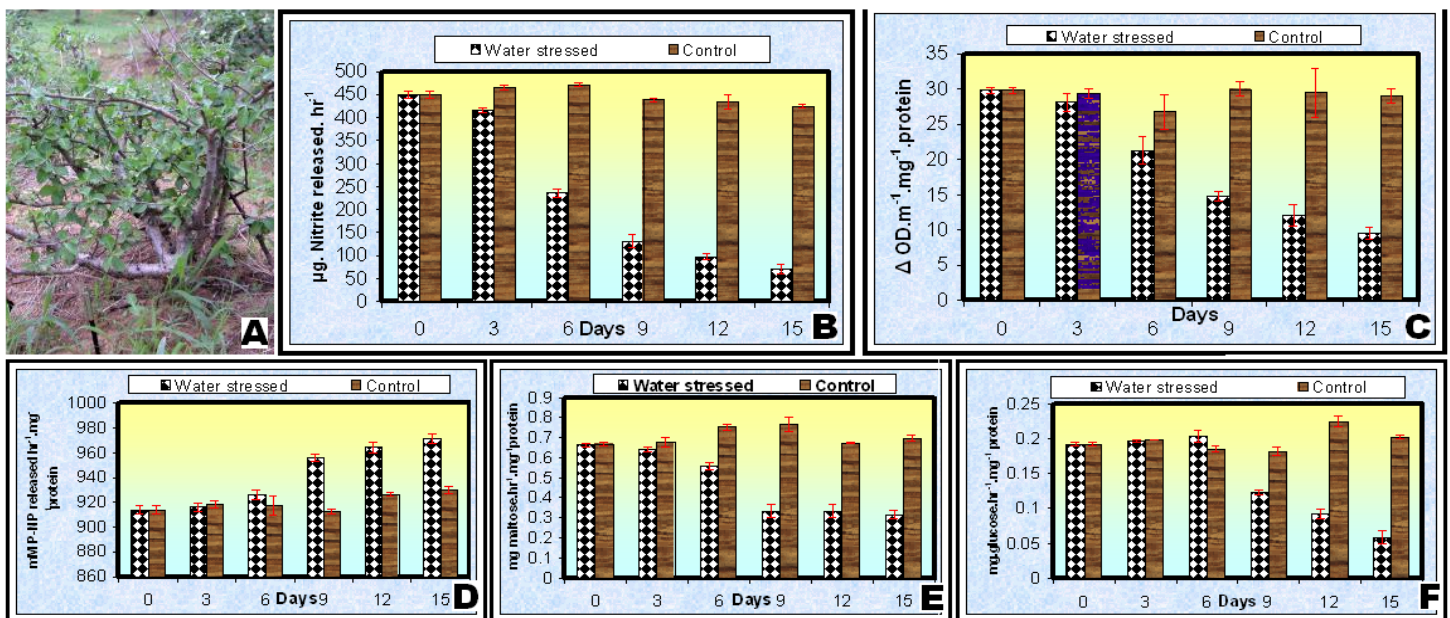


Fig. 1: (A) Mature plant of *C. wightii*; Drought induced changes in nitrate reductase, (B) Peroxidase,

(C) Acid phosphatase, (D) α -amylase, (E) Invertase, (F) *C. wightii* under drought stress

Activity of α -amylase declined marginally till day 6 and declined sharply thereafter in the guggul plants subjected to water stress. The α -amylase activity in non-stressed plants increased marginally during first nine days (Fig. 1 E). Similarly results were also observed in pearl millet [22]. Invertase activity initially increased till day 6 and then declined sharply with an increasing water starvation (Fig. 1 F). The initial increase in invertase activity in plants indicates its involvement in water stress tolerance mechanism. Thind and Malik [23] also observed an increase in acid invertase activity in wheat at low stress levels.

CONCLUSIONS

In the present investigations, biochemical basis of drought tolerance in the succulent plant *C. wightii* was studied. Biochemical analysis indicates that plants are quite resistant and well developed to endure drought stress by modulating the key enzymes such as enzymes nitrate reductase, peroxidase, acid phosphatase, α -amylase and invertase. The study will be highly beneficial in the development of transgenic plants against drought stress.

ACKNOWLEDGMENT

The authors are thankful to Prof. C. P. Malik, Prof. Reto Strasser and Prof. B. Robert for constant blessing and academic encouragement.

REFERENCES

- [1] Araus JL, Slafer GA, Reynolds MP, Royo C. Plant breeding and drought in C3 cereals: what should we breed for? *Ann. Bot.*, 2002; 89: 925-40.
- [2] Molnár I, Gáspár L, Sárvári É, Dulai S, Hoffmann B, et al. Physiological and morphological responses to water stress in *Aegilops biuncialis* and *Triticum aestivum* genotypes with differing tolerance to drought. *Funct. Plant Biol.*, 2004; 31: 1149-59.
- [3] Molnar I, Dulai S, Csernak Á, Pronay J, Molnar-Lang M. Photosynthetic responses to drought stress in different *Aegilops* species. *Acta Biol. Szeged.*, 2005; 49(1-2): 141-42.
- [4] Bohnert HJ, Nelson DE, Jensen RG. Adaptations to environmental stresses. *Plant Cell*, 1995; 7: 1099-111.
- [5] Jaworski E. Nitrate reductase assay in intact plant tissues. *Biochem. Biophys. Res. Commun.*, 1971; 43: 1247-79.
- [6] Racusen D, Foote M. Protein synthesis in dark grown bean leaves. *Can. J. Bot.*, 1965; 43: 817-24.
- [7] Zink MW, Veliky IA. Acid phosphatase of *Ipomoea* spp. cultured *in vitro*. 1. Influence of pH and inorganic phosphate on the formation of phosphatases. *Can. J. Bot.*, 1979; 57: 739-53.
- [8] Bernfeld P. Amylases α and β . *Methods in Enzymol.*, 1955; 1: 149-58.
- [9] Harris GP, Jeffcoat B. Effects of temperature on the distribution of ^{14}C -labelled assimilate in the flowering shoot of carnation. *Ann. Bot.*, 1974; 38: 77-83.

- [10] Bray EA. Plant responses to water deficit. *Trends Plant Sci.*, 1997; 2: 48-54.
- [11] Tabaeizadeh Z. Drought induced responses in plant cells. *Int Rev Cyto*, 1998; 182: 193-247.
- [12] Grumet R, Hanson AD. Genetic evidence for an osmoregulatory function of glycine betanin accumulation in barley. *Aus. J. Plant Physiol.*, 1986; 13: 353-64.
- [13] Garg BK, Kathju S, Lahiri AN, Vyas SP. Drought resistance in pearl millet. *Biologia. Plantarum.*, 1981; 23: 182-85.
- [14] Saxena HK, Yadav RS, Parihar SKS, Singh HB, Singh GS. Susceptibility and recovery potential of rice genotypes to drought at different growth stages. *Ind. J. Plant Physiol.*, 1996; 1: 198-202.
- [15] Morilla CA, Boyer JS, Hageman RL. Nitrate reductase activity and polyribosomal content of corn (*Zea mays* L.) having low leaf water potentials. *Plant Physiol.*, 1973; 51: 817-24.
- [16] Rajagopal V, Balasubramanian V, Sinha K. Diurnal fluctuations in relative water content, nitrate reductase and proline content in water stressed and non-stressed wheat. *Physiol. Plant*, 1977; 40: 69-71.
- [17] Teare ID, Manam R, Kanemasu ET. Diurnal and seasonal trends in nitrate reductase activity in field grown *Sorghum* plants. *J. Agron.*, 1974; 66: 733-36.
- [18] Alekseeva AY, Ramazanov LK. Catalytic functions of chloroplast peroxidase under dehydration conditions. *Izv. Akad. Ser. Biol. Navk. (SSR)*, 1973; 6: 903-05.
- [19] Chiung-Yueh S, Ching-Hvei K. Induction of acid phosphatase in detached rice leaves under stress conditions. *Bot. Bull. Head Sin.*, 1998; 39: 29-32.
- [20] Sharma AD, Thakur M, Rana M, Singh K. Effect of plant growth hormones and abiotic stresses on germination, growth and phosphatase activities in *Sorghum bicolor* (L.) Moench seeds. *African J. Biotech.*, 2004; 3(6): 308-12.
- [21] Olmos E, Hellin E. Cytochemical localization of ATPase plasma membrane and acid phosphatase by cerium based in a salt-adapted cell line of *Pisum sativum*. *J. Exp. Bot.*, 1997; 48: 1529-35.
- [22] Garg BK, Vyas SP, Kathju S, Lahiri AN. Influence of water deficit stress at various growth stages on some enzymes of nitrogen metabolism and yield in cluster bean genotypes. *Ind. J. Plant Physiol.*, 1998; 3(3): 214-18.
- [23] Thind SK, Malik CP. Role of acid invertase and glycosidases in growth of wheat seedlings subjected variable water stress levels. *Ind. J. Exp. Biol.*, 1989; 27: 279-89.

International Journal of Life-Sciences Scientific Research (IJLSSR) Open Access Policy

Authors/Contributors are responsible for originality, contents, correct references, and ethical issues.

IJLSSR publishes all articles under Creative Commons Attribution- Non-Commercial 4.0 International License (CC BY-NC).

<https://creativecommons.org/licenses/by-nc/4.0/legalcode>



How to cite this article:

Soni V, Swarnkar PL: Drought Induced Biochemical Changes in *Commiphora wightii*. *Int. J. Life Sci. Scienti. Res.*, 2017; 3(4):1247-1249. DOI:10.21276/ijlssr.2017.3.4.24

Source of Financial Support: Nil. Conflict of interest: Nil